





Biosafety Clearing-House (BCH)

LIVING MODIFIED ORGANISM (LMO)

BCH-LMO-SCBD-116256-2

? Decisions on the LMO ? Risk Assessments

LAST UPDATED: 01 OCT 2021

Living Modified Organism identity

The image below identifies the LMO through its unique identifier, trade name and a link to this page of the BCH. Click on it to download a larger image on your computer. For help on how to use it go to the LMO quick-links page.

https://bch.cbd.int/database/record?documentID=116256



MON-87427-7 \times MON-8746Ø-4 Drought-tolerant maize with tissue-specific herbicide tolerance



Read barcode or type above URL into internet browser to access information on this LMO in the Biosafety Clearing-House @ 5CBD 2012

Name

Drought-tolerant maize with tissue-specific herbicide tolerance

ΕN

Transformation event

MON 87427 × MON 87460

Unique identifier

MON-87427-7 × MON-8746Ø-4

Developer(s)

- PERSON: BAYER CROPSCIENCE | BCH-CON-SCBD-111462-3

PERSON

Bayer CropScience

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RELATED ORGANIZATION

Description

The maize (*Zea mays*) was modified for tissue-specific herbicide tolerance and drought tolerance through cross breeding of modified parental lines. For glyphosate tolerance, the maize expresses *Agrobacterium tumefaciens* 5-enolpyruvylshikimate-3-phosphate synthase

ΕN

(*cp4-epsps*), which is a bacterial variant of the endogenous gene involved in the biosynthesis of aromatic amino acids (shikimate pathway). The bacterial protein does not bind the herbicide with high affinity and thus allows for the continued biosynthesis of essential aromatic amino acids. Due to the combination of promoter and intron genetic elements (see 'Notes regarding the specific genetic elements' below), *cp4-epsps* is expressed in vegetative and female-specific tissues, thus allowing for glyphosate-treated maize to serve as the female parent during the production of hybrid seed. For drought tolerance, the maize expresses *Bacillus subtilis* cold shock protein, which binds RNA and maintains cellular functions under water-limited conditions (improvement of natural abiotic stress responses, also to cold stress). The maize also contains an *Escherichia coli* neomycin phosphotransferase II cassette, which allowed for kanamycin selection during transformation of a parental line.

Recipient Organism or Parental Organisms

The term "Recipient organism" refers to an organism (either already modified or non-modified) that was subjected to genetic modification, whereas "Parental organisms" refers to those that were involved in cross breeding or cell fusion.

BCH-ORGA-SCBD-246-6 ORGANISM | ZEA MAYS (MAIZE, CORN, MAIZE)

Crops

BCH-LMO-SCBD-104758-3 LIVING MODIFIED ORGANISM | MON-87427-7 - MAIZE MODIFIED FOR TISSUE SELECTIVE GLYPHOSATE TOLERANCE |

Resistance to herbicides - Glyphosate

BCH-LMO-SCBD-103066-6 LIVING MODIFIED ORGANISM | MON-8746Ø-4 - DROUGHTGARD™ MAIZE

Resistance to antibiotics - Kanamycin Tolerance to abiotic stress - Cold / Heat, Drought

Characteristics of the modification process

Vector

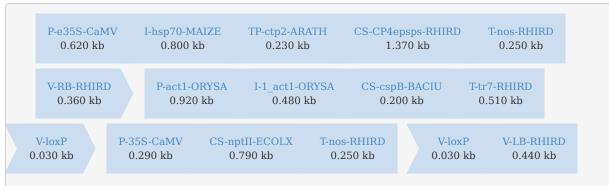
PV-ZMAP1043; PV-ZMAP595

ΕN

Techniques used for the modification

Cross breeding

Genetic elements construct



Introduced or modified genetic element(s)

Some of these genetic elements may be present as fragments or truncated forms. Please see notes below, where applicable.

BCH-GENE-SCBD-100366-6 CAMV ENHANCED 35S PROMOTER

Promoter

BCH-GENE-SCBD-100359-7 HSP70 INTRON | (MAIZE, CORN)

Intron

BCH-GENE-SCBD-100365-6 CHLOROPLAST TRANSIT PEPTIDE 2 | (THALE CRESS)

Transit signal

BCH-GENE-SCBD-14979-7 5-ENOLPYRUVYLSHIKIMATE-3-PHOSPHATE SYNTHASE GENE

Protein coding sequence | Resistance to herbicides (Glyphosate)

BCH-GENE-SCBD-100269-8 NOPALINE SYNTHASE GENE TERMINATOR

Terminator

BCH-GENE-SCBD-101415-9 TI PLASMID LEFT BORDER REPEAT

Plasmid vector

BCH-GENE-SCBD-100364-5 RICE ACTIN 1 GENE PROMOTER | (RICE)

Promoter

BCH-GENE-SCBD-100355-6 RICE ACTIN 1, INTRON | (RICE)

Intron

BCH-GENE-SCBD-103065-7 COLD SHOCK PROTEIN GENE

Protein coding sequence | Tolerance to abiotic stress (Cold / Heat, Drought)

BCH-GENE-SCBD-103067-9 TRANSCRIPT 7 GENE 3' UNTRANSLATED REGION

Terminator

BCH-GENE-SCBD-103069-3 LOXP RECOMBINATION SITE

recombination site

BCH-GENE-SCBD-100287-7 CAMV 35S PROMOTER

Promoter

BCH-GENE-SCBD-15001-5 NEOMYCIN PHOSPHOTRANSFERASE II | (BACTERIA)

Protein coding sequence | Resistance to antibiotics (Kanamycin)

BCH-GENE-SCBD-101416-6 TI PLASMID RIGHT BORDER REPEAT

Plasmid vector

Notes regarding the genetic elements present in this LMO

DNA insert from MON87427 PV-ZMAP1043

Transcription of 5-enolpyruvylshikimate-3-phosphate synthase (*cp4-epsps*) from *Agrobacterium tumefaciens* commences from the *Cauliflower mosaic virus* (CaMV) enhanced 35S promoter and ends at the *A. tumefaciens* nopaline synthase (*nos*) gene terminator. The transcript contains a *Zea mays* heat shock protein 70 (*hsp70*) intron, *Arabidopsis thaliana* N-terminal chloroplast transit peptide sequence, and *cp4-epsps*. The CaMV enhanced 35S promoter-*hsp70* combination promotes gene expression in female and vegetative tissues, but not in male reproductive tissues (pollen microspores and tapetum).

ΕN

Note:

- Southern blot analyses indicate that a single copy of the T-DNA was inserted at a single site in the parental maize genome and no plasmid vector backbone sequences were detected to have been integrated. DNA sequencing analyses further indicated that the expected T-DNA sequences were integrated.

-The *cp4-epsps* coding sequence is the codon optimized coding sequence of the *aroA* gene from *Agrobacterium sp.* strain CP4 encoding CP4 EPSPS.

DNA insert from MON87460 vector PV-ZMAP595

The T-DNA insert contains the following gene cassettes: *Bacillius subtillus* cold shock protein (*cspB*) and *Escherichia coli* neomycin phosphotransferase II (*nptII*).

Transcription of *cspB* is under control of the *Oryza sativa* actin 1 promoter and *Agrobacterium tumefaciens* transcript 7 gene 3' untranslated region. The transcript initially contains an *O. sativa* actin 1 intron for enhanced gene expression of *cspB*. The sequence is removed (spliced) prior to protein translation. Constitutive expression of *cspB* is expected due to the actin promoter.

Transcription of *nptll* is under control of the *Cauliflower mosaic virus* (CaMV) 35S promoter and *A. tumefaciens* nopaline synthase terminator. High levels of transcription are expected due to the CaMV promoter.

Note:

- The coding sequence of *cspB* has been codon optimized for optimal expression within plant
- Southern blot analysis indicated that no vector backbone sequences were inserted into the parental genome
- Southern blot analysis indicated that the parental genome contains a single insertion
- Sequencing analyses confirm the Southern blot analyses.
- A 22 base pair deletion of genomic DNA at the insert-to-plant DNA junction occurred.
- *loxP* sites can be found in the parental genome and could potentially allow for the excision of the *nptll* cassette by CRE recombinase.

For more information, kindly refer to the parental LMO records.

LMO characteristics

Modified traits

Resistance to herbicides

Glyphosate

Resistance to antibiotics

Kanamycin

Tolerance to abiotic stress

Cold / Heat

Drought

Selectable marker genes and reporter genes

Common use(s) of the LMO

Food

Feed

Detection method(s)

External link(s)

? MON-87427-7 - EU Reference Laboratory for GM Food and Feed (EURL-GMFF) (English)

? MON-8746Ø-4 - EU Reference Laboratory for GM Food and Feed (EURL-GMFF) (English)

? MON-87427-7 - GMO Detection method Database (GMDD) (English)

? MON-8746Ø-4 - GMO Detection method Database (GMDD) (English)

Additional Information

Other relevant website addresses and/or attached documents

? EUginius - MON-87427-7 x MON-8746Ø-4 (*English*)

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Further Information

Questions about the Cartagena Protocol on Biosafety or the operation of the Biosafety Clearing-House may be directed to the Secretariat of the Convention on Biological Diversity.

Secretariat of the Convention on Biological Diversity

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